

Perspective: L-arginine and L-citrulline Supplementation in Pregnancy: A Potential Strategy to Improve Birth Outcomes in Low-Resource Settings

Andrea M Weckman, 1,2 Chloe R McDonald, 2 Jo-Anna B Baxter, 3,4 Wafaie W Fawzi, 5 Andrea L Conroy, 2 and Kevin C Kain 1,6,2

¹Laboratory Medicine and Pathobiology, ²Sandra A Rotman Laboratories, Sandra Rotman Centre for Global Health, University Health Network-Toronto General Hospital, Toronto, Canada; ³Department of Nutritional Sciences, and ⁴Centre for Global Child Health, Hospital for Sick Children, Toronto, Canada; ⁵Department of Global Health and Population, Harvard TH Chan School of Public Health, Boston, MA; and ⁶Tropical Disease Unit, Division of Infectious Diseases, Department of Medicine, University of Toronto, Toronto, Canada

ABSTRACT

The available data support the hypothesis that L-arginine or L-citrulline supplementation would be suitable for implementation in resource-constrained settings and will enhance placental vascular development and improve birth outcomes. In resource-constrained settings, the rates of adverse birth outcomes, including fetal growth restriction, preterm birth, and low birth weight, are disproportionately high. Complications resulting from preterm birth are now the leading cause of mortality in children <5 y of age worldwide. Despite the global health burden of adverse birth outcomes, few effective interventions are currently available and new strategies are urgently needed, especially for low-resource settings. L-arginine is a nutritionally essential amino acid in pregnancy and an immediate precursor of nitric oxide. During pregnancy, placental and embryonic growth increases the demand for L-arginine, which can exceed endogenous synthesis of L-arginine from L-citrulline, necessitating increased dietary intake. In many low-resource settings, dietary intake of L-arginine in pregnancy is inadequate owing to widespread protein malnutrition and depletion of endogenous L-arginine due to maternal infections, in particular malaria. Here we examine the role of the L-arginine-nitric oxide biosynthetic pathway in pregnancy including placental vascular development and fetal growth. We review the evidence for the relations between altered L-arginine bioavailability and pregnancy outcomes, and strategies for arginine supplementation in pregnancy. Existing studies of L-arginine supplementation in pregnancy in high-resource settings have shown improved maternal and fetal hemodynamics, prevention of pre-eclampsia, and improved birth outcomes including higher birth weight and longer gestation. Arginine supplementation studies now need to be extended to pregnant women in low-resource settings, especially those at risk of malaria. Adv Nutr 2019;10:765–777.

Keywords: L-arginine, L-citrulline, amino acid, pregnancy, birth outcomes, maternal malnutrition, supplementation, placental vasculature, nitric oxide, low- and middle-income countries

Adverse Birth Outcomes Are Disproportionately High in Low-Resource Settings

Complications resulting from preterm birth (PTB; delivery before 37 weeks of gestation) are the leading cause of death in children under the age of 5 y (1, 2). An estimated 15 million preterm deliveries occur each year and 1 million preterm infants die in their first month of life owing to complications of prematurity including birth asphyxia, hypothermia, infection, and respiratory distress syndrome (3). A further 800,000 infant deaths every year are attributed to fetal growth restriction (FGR) (4). Among newborns that survive, PTB and FGR are associated with poor long-term development including childhood stunting, neurological

impairments, and an increased risk of chronic disease (4–6). Despite the growing global burden of PTB and other adverse birth outcomes, there are a limited number of safe and effective intervention strategies to improve health outcomes in pregnancy.

The likelihood of delivering preterm is greatly increased in resource-constrained settings. More than 60% of all PTBs and three-quarters of neonatal deaths resulting from PTB complications occur in South Asia and sub-Saharan Africa (1). The disparity in adverse birth outcomes between high-income countries and low- and middle-income countries (LMICs) is due in large part to modifiable risk factors including infectious disease (e.g., malaria, HIV, and sexually

transmitted diseases), anemia, undiagnosed pre-eclampsia, and maternal malnutrition (7).

Maternal Malnutrition and Infection Are Key Contributors to Adverse Birth Outcomes in Resource-Constrained Settings

Malnutrition is a key contributor to global maternal morbidity and mortality, adverse birth outcomes, and longterm health consequences for the child (8-10). In 2016, an estimated 815 million individuals worldwide were chronically malnourished, with the vast majority living in sub-Saharan Africa and South Asia (11). Protein deficiency and poor-quality protein consumption (e.g., cereals lacking essential amino acids) are widespread across sub-Saharan Africa (12, 13) and low circulating concentrations of essential and conditionally essential amino acids including L-arginine are associated with impaired child growth (13). Infants, children, and women of reproductive age are at particular risk of nutritional deficiencies given their increased requirements (8). Over 10% of reproductive-aged women in LMICs are underweight (BMI $<18.5 \text{ kg/m}^2$) (4, 14, 15). Poor nutritional status, reflected by low BMI, insufficient weight gain across pregnancy, and micro- and macronutrient deficiencies, contributes to increased risk of PTB, FGR, and neurodevelopmental deficits in children (8, 9, 16, 17). Protein supplementation has been shown to reduce rates of adverse pregnancy outcomes. Systematic reviews of intervention studies using balanced-energy protein in pregnancy have reported significant reductions in the incidence of small-forgestational-age outcomes and stillbirth (18, 19). However, high rates of adverse birth outcomes still occur and more targeted strategies (e.g., L-arginine supplementation) may be required to further reduce poor birth outcomes, especially those related to common infections in pregnancy (e.g., malaria and HIV).

In this Perspective, we examine malaria in pregnancy as a model of infections whose negative impact on birth outcomes could be mitigated by arginine supplementation. Worldwide, 125 million pregnancies are at risk of malaria infection each year (20). Malaria infections are more frequent and severe during pregnancy and result in an estimated

Supported by Global Alliance to Prevent Prematurity and Stillbirth and Grand Challenges in Global Health: Preventing Preterm Birth Initiative grant no. 12003, Healthy Birth Initiative grant OPP1033514 (to KCK); a Grand Challenges Canada Rising Stars in Global Health grant (to ALC); Canadian Institutes of Health Research (CIHR) Foundation grant FDN-148139 (to KCK); a Canada Research Chair (to KCK); and CIHR Doctoral Award GSD-157907 (to AMW). Author disclosures: AMW, CRM, J-ABB, WWF, ALC, and KCK, no conflicts of interest. AMW and CRM contributed equally to this work.

Perspective articles allow authors to take a position on a topic of current major importance or controversy in the field of nutrition. As such, these articles could include statements based on author opinions or point of view. Opinions expressed in Perspective articles are those of the author and are not attributable to the funder(s) or the sponsor(s) or the publisher, Editor, or Editorial Board of Advances in Nutrition. Individuals with different positions on the topic of a Perspective are invited to submit their comments in the form of a Perspectives article or in a Letter to the Editor

Address correspondence to KCK (e-mail: kevin.kain@uhn.ca).

Abbreviations used: ADMA, asymmetric dimethylarginine; eNOS, endothelial nitric oxide synthase; FGR, fetal growth restriction; LMIC, low- and middle-income country; NOS, nitric oxide synthase; PIGF, placental growth factor; PTB, preterm birth; RCT, randomized controlled trial; sFlt1, soluble fms-like tyrosine kinase-1; sGC, soluble guanylate cyclase; VEGF, vascular endothelial growth factor.

750,000 low-birth-weight deliveries (due to PTB and/or FGR) in sub-Saharan Africa alone (21, 22). Considerable evidence supports an interaction between malaria in pregnancy and maternal malnutrition in worsening pregnancy outcomes in malaria-endemic regions (23). A recent metaanalysis reported an increased risk of low birth weight in women who were both malnourished and exposed to malaria, compared with women affected by either risk factor alone (24). Identification of common pathways leading to adverse birth outcomes affected by both malaria infection and malnutrition would enable more targeted and effective prevention strategies. Here we examine the role of one such common pathway, the L-arginine-nitric oxide (NO) biosynthetic pathway. The L-arginine-NO pathway is important for healthy pregnancies, dependent on maternal nutritional status, and negatively affected by maternal infections including malaria. We hypothesize that a combination L-arginine and L-citrulline supplement for pregnant women in LMICs at risk of malaria infection could increase bioavailable NO, enhance placental vascular development, and improve birth outcomes.

L-arginine-NO Biogenesis Plays a Critical Role in Placental Vascular Development

The formation of a functional and adequately vascularized placenta is essential for a healthy pregnancy and birth outcome (25). The establishment of vascularized placental tertiary villi, the site of nutrient exchange between maternal and fetal blood, depends on the tightly regulated processes of vasculogenesis, or the de novo formation of blood vessels, and angiogenesis, the remodeling of existing vasculature (26, 27).

A growing body of evidence links placental vascular pathology with poor fetal growth and adverse birth outcomes (28-30). Altered uterine (maternal side) and umbilical (fetal side) hemodynamics resulting from disruptions to normal placental vasculogenesis or angiogenesis has been mechanistically linked to adverse outcomes including fetal hypoxia, hypoglycemia, and impaired fetal organ growth (31, 32). Moreover, insufficient vascular adaptation to accommodate changing hemodynamic activity as the fetus grows can contribute to maternal gestational diabetes, pre-eclampsia, and adverse birth outcomes such as PTB and FGR (33, 34).

L-arginine is involved in multiple pathways that contribute to healthy placental and fetal function and development. L-arginine is the physiologically active isomer of arginine (compared with the D-isomer) and thus the supplementation substrate; therefore, we refer to L-arginine as "arginine" in this article. The body can produce arginine via de novo synthesis from L-citrulline, as well as during protein turnover; therefore, exogenous arginine is not the sole source. De novo synthesis of arginine accounts for 5-15% of circulating plasma arginine (35). Endogenous arginine synthesis involves a diverse set of enzymes that are differentially expressed across organs and cell types. Adult mammals synthesize most de novo arginine in the kidney; however, the location of synthesis and its relative contribution to circulating arginine depend on species and stage of development (35). Net production of arginine in a given organ is determined by the relative expression of arginine synthesis and arginine catabolic enzymes (35).

Arginine is a precursor in numerous biological pathways including protein synthesis and the production of ornithine and urea, glutamate, NO, creatine, proline, and polyamines (35, 36). The complex metabolic pathways of arginine have been well characterized (35). A number of enzymes and competitive inhibitors including arginase and asymmetric dimethylarginine (ADMA) influence the equilibrium of arginine use between metabolic pathways, dictating the bioavailability of NO (Figure 1). Three principal isoforms of nitric oxide synthase (NOS) enzymes produce NO from arginine: neuronal NOS (nNOS or NOS1) and endothelial NOS (eNOS or NOS3) which are both constitutively expressed, and inducible NOS (iNOS or NOS2) induced in response to inflammation. The enzyme arginase competes with NOS for arginine, promoting the production of ornithine and urea rather than NO. ADMA competitively inhibits NOS, whereas symmetric dimethylarginine enhances oxidative stress and, at high concentrations, impairs arginine uptake by cells, directly and indirectly preventing arginine conversion to NO (37).

The arginine-NO pathway is an important regulator of vascular development, and altered arginine and NO

bioavailability may impair vasculogenesis and angiogenesis and negatively affect fetal growth and survival (38). NO is a gaseous signaling molecule initially identified for its vasodilatory activity, acting via soluble guanylate cyclase (sGC) in smooth muscle cells to control vascular tone. It has since been shown to have critical roles in regulation of neurotransmission, angiogenesis, inflammation, and cell adhesion by mechanisms including activation of sGC and by s-nitrosylation of numerous regulatory proteins (39, 40). At the endothelial interface, NO regulates critical mediators of angiogenesis including the vascular endothelial growth factor (VEGF) [e.g., placental growth factor (PIGF) and its receptor soluble fms-like tyrosine kinase-1 (sFlt1)] and angiopoietin-tunica interna endothelial cell kinase 2 (Tie-2) (e.g., angiopoietin-1 and angiopoietin-2 and their receptor soluble Tie-2) protein families. The induction of angiogenesis by angiopoietin and VEGF signaling is dependent on protein kinase-mediated downstream activation of eNOS in the endothelium (and in pregnancy, placental cytotrophoblasts and syncytiotrophoblasts), and consequent production of NO from arginine (30, 38, 41, 42). Collectively the available evidence implicates dysregulation of arginine-NO biogenesis in a range of vascular pathologies including cardiovascular disease and pre-eclampsia (43, 44).

L-citrulline is produced as a by-product of NO synthesis from arginine and can be recycled in a feedback loop to

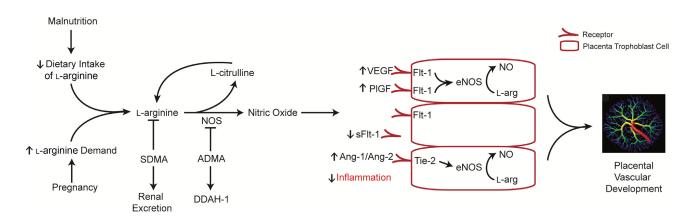


FIGURE 1 The L-arginine-NO biosynthetic pathway regulates key vasculogenic and angiogenic factors in pregnancy. Demand for L-arginine is increased during pregnancy to support the rapidly growing placenta and fetus. L-arginine is acquired in protein-rich foods, which are often lacking in diets in low-resource settings. Therefore, reduced dietary intake, in combination with increased demand in pregnancy, may place pregnant women at risk of arginine deficiency and reduced bioavailable NO. L-arginine is converted to NO via NOS enzymes. L-citrulline, which is also acquired through dietary intake, is produced as a by-product of NO synthesis from L-arginine, and can be recycled in a feedback loop to generate more arginine. Several enzymes and competitive inhibitors such as arginase, ADMA, and SDMA influence the equilibrium of arginine use between metabolic pathways and affect the bioavailability of NO. ADMA competitively inhibits NOS, whereas SDMA can inhibit the cellular uptake of L-arginine, directly and indirectly preventing L-arginine conversion to NO. The enzyme DDAH degrades methylarginines, including ADMA. At the placental trophoblast, NO interacts with critical mediators of angiogenesis (PIGF, VEGF, the angiopoietins). The induction of angiogenesis by angiopoietin and VEGF proteins is dependent on downstream activation of eNOS and production of NO from L-arginine. NO signaling also acts in a positive feedback loop to increase endothelial amounts of VEGF and Ang-1. Furthermore, NO reduces inflammation-mediated endothelial activation by reducing expression of vascular adhesion molecules and proinflammatory cytokines. ADMA, asymmetric dimethylarginine; Ang-1/-2, angiopoietin-1 and -2; DDAH-1, dimethylarginine dimethylaminohydrolase-1; eNOS, endothelial nitric oxide synthase; NOS, nitric oxide synthase; PIGF, placental growth factor; SDMA, symmetric dimethylarginine; sFlt-1, soluble fms-like tyrosine kinase-1; Tie-2, tunica interna endothelial cell kinase 2; VEGF, vascular endothelial growth factor.

generate more arginine. Citrulline is also acquired via dietary intake and through synthesis from glutamine, glutamate, and proline in the intestine (35). The conversion of citrulline to arginine in the intestinal-renal axis accounts for most de novo arginine production (35, 45). Disease states with reduced L-citrulline production (i.e., sepsis) exhibit decreased bioavailable arginine and NO, contributing to pathology (46). During conditions of decreased dietary arginine intake, as in many resource-constrained settings (where Lcitrulline intake would also be low), de novo endogenous synthesis of arginine from L-citrulline is not enhanced to compensate (47). Therefore, bioavailable arginine necessary for optimal physiologic function may be outstripped in conditions of simultaneously increased metabolic demand (e.g., pregnancy, infection) and decreased dietary arginine and citrulline.

In addition to NO, several other arginine metabolites including polyamines, creatine, and proline are also necessary for healthy pregnancies (48, 49). Creatine is required for fetal neurological development and muscle function, whereas proline is an important amino acid for cartilage development (48, 49). Polyamines interact with RNA and DNA to modulate protein synthesis and cell growth and function (50). Polyamines are synthesized from L-arginine via the agmatine and ornithine metabolic pathways and are required through several stages of pregnancy, including implantation, early embryogenesis, fetal growth, and placental development (51, 52). Pharmacological and diet-driven reductions in polyamine bioavailability in pregnant rodents and large mammals have been associated with abnormal placentation, FGR, and pregnancy loss (51, 52).

NO also plays several regulatory roles throughout placental development, beginning with its role as a facilitator of implantation and endovascular trophoblast invasion (30, 38, 53, 54). The processes of trophoblast cell migration and invasion, vessel development, and vessel maturation in the placenta are facilitated by the VEGF and angiopoietin pathways, which induce NO production in the endothelium (26, 30, 38, 55, 56). Early in pregnancy, eNOS is also expressed in several placental cell types including cytotrophoblasts, extravillous trophoblasts, and syncytiotrophoblasts (38). As pregnancy progresses, placental eNOS expression increases and becomes localized primarily to syncytiotrophoblasts and vascular endothelial cells (42, 57). In the absence of eNOS, VEGF does not exhibit its regular angiogenic effects (58). NO also promotes a proangiogenic environment by increasing trophoblast expression of PIGF and VEGF and decreasing expression of the antiangiogenic factor sFlt1 (59). Recent evidence has implicated changes in angiogenic factors mediated by NO signaling (e.g., PIGF, VEGF, sFlt1, angiopoietin-1/2), as well as NO itself, in placental insufficiency and adverse pregnancy outcomes including gestational hypertension, pre-eclampsia, PTB, and FGR (34, 60-69). Factors that decrease the ability of eNOS to convert arginine to NO (i.e., increased ADMA or decreased bioavailable arginine) have also been implicated in the pathobiology of pregnancy disorders including FGR and pre-eclampsia (70–73). After

placental vasculature is established, NO acts as a vasodilator by activating sGC in vascular smooth muscle cells to induce relaxation. NO-mediated vasodilation is necessary during pregnancy to modulate maternal blood pressure, systemic blood flow, and adequate placental perfusion to support fetal growth. NO synthesis is required to decrease the risk of pregnancy-related hypertensive disorders, including preeclampsia and eclampsia.

Collectively these data indicate there is an increased demand for bioavailable NO, polyamines, and arginine during pregnancy in order to support placental and fetal growth and function. In humans, there is a physiological increase in arginine synthesis during the second trimester, a decrease in circulating ADMA, and a corresponding increase in NO synthesis (74, 75). Despite increased in vivo production, however, pregnancy has been described as a state of arginine deficiency because endogenous production cannot meet the increased demand from placental and fetal growth (76). Thus, exogenous intake of arginine is particularly important for pregnant women and dietary supplementation may be required in situations where dietary intake is inadequate.

Arginine Nutritional Requirements, Dietary Sources, and Supplementation in Pregnancy

Protein-rich foods such as meats, dairy products, nuts, seeds, soy, and pulses are considered to be rich in arginine. The relative amount of arginine in various proteins ranges from 3% to 15% (77, 78). Most cereals are deficient in arginine, with only 3–4% of their low protein content derived from arginine (78). Vegetables and fruits contain a negligible amount of protein and thus contain little arginine. As a result, variation in dietary patterns could account for the differences in plasma arginine observed globally (78). In resource-constrained settings, like sub-Saharan Africa, only 9% of dietary energy comes from arginine-rich foods such as meats, pulses, and seeds, whereas >60% comes from arginine-poor roots, tubers, and cereal-based staples (12). Therefore, patterns of dietary intake indicate the potential for widespread arginine deficiency in these areas, especially in pregnant women. Arginine is now considered to be nutritionally essential for optimal health, fertility (i.e., spermatogenesis in human males), and reproduction (79). However, because arginine has traditionally been considered a non- or conditionally essential amino acid in the context of maintaining nitrogen balance (79), comprehensive reference materials for dietary arginine requirements have not been compiled.

Normal dietary intake in Western countries is \sim 4–5 g/d (36). The tolerable upper intake limit of arginine supplementation, especially over long periods, has not been definitively set (80). Studies in pigs and rats have not reported adverse effects with supplemental doses \leq 630 mg arginine \cdot kg body weight⁻¹ \cdot d⁻¹ and 3.6 g arginine \cdot kg body weight⁻¹ \cdot d⁻¹, respectively, for 91 d (81). Using conversion factors provided by the FDA, this translates to an estimated

40 g/d for a 70-kg human adult (81, 82). Human trials of arginine supplementation in adults have been inconsistent in dose, duration, and sample population; however, the absence of reported adverse events remains consistent (81, 83). The observed safe amount for arginine supplementation in adults is currently 30 g/d for 90 d (84). Mild gastrointestinal side effects have been reported after large amounts of arginine in a single dose, but using multiple smaller doses is reported to reduce gastrointestinal intolerance (84).

The required dose of arginine during pregnancy has not been determined. However, the daily average intake of arginine is 4.3 g/d among pregnant women in the United States, a country where arginine intake is considered to be sufficient (85). Arginine consumption has been estimated at \leq 2–3 g/d in low-resource settings, well below the intake of women in resource-rich settings (12). Pregnancy is a state of relative arginine deficiency and infections such as malaria further deplete arginine (86); therefore, the low dietary intake of arginine in resource-constrained, malaria-endemic settings may put pregnant women at risk of hypoarginemia. The critical role that the arginine–NO biosynthetic pathway plays in regulating vascular development and other pathways important for healthy pregnancy outcomes suggests that supplemental arginine in LMICs may reduce adverse birth

A supplement-based strategy is generally identified as the most effective approach for populations that have a high prevalence of nutrient deficiency and for target groups where a required nutrient is difficult to attain through normal diet (e.g., LMICs) (87). Possible supplementation platforms for arginine include syrups, food bars, tablets (chewable or effervescent), caplets, and powders (including crystalline forms of arginine like arginine-HCl and arginine α -ketoglutarate). Supplementation of arginine and Lcitrulline in ready-to-use supplementary food in Tanzanian children with sickle-cell disease reported no issues with tolerability and a compliance rate of 95% (88), suggesting this may be an effective formulation for arginine supplementation in LMICs.

One challenge in arginine supplementation is arginase, which catalyzes the metabolism of arginine into urea and is highly active in the intestine of adults. Only 60% of oral arginine evades intestinal metabolism (47) and another 15% of circulating arginine is metabolized in the liver (89). An approach to circumvent arginine metabolism is the administration of L-citrulline, which can be converted into arginine but is not a substrate for arginase (90). Thus, coadministration of L-citrulline and arginine could bypass this barrier and increase the efficacy of arginine supplementation. Recent studies support the effectiveness of oral Lcitrulline in increasing bioavailable arginine concentrations and NO-dependent signaling (91, 92). The combination of oral L-citrulline and arginine supplementation in adult males increased plasma arginine more effectively than did either supplement alone (93). These studies support the hypothesis that supplementation with arginine plus

L-citrulline would be the most effective strategy to increase bioavailable arginine and NO and improve birth outcomes.

Studies of Arginine Supplementation in Pregnancy

A large body of work has investigated arginine supplementation in pregnancy in mammals (49, 94). Supplementation with arginine and other members of arginine metabolic pathways (e.g., glutamine) results in increased placental growth (95, 96), fetal viability (95, 97-99), litter size (95, 96, 100), and birth weight (49, 95-100) in pigs and sheep. Animal models using low-protein diet-induced FGR have reported increases in NO production, fetal:placental weight ratio, and fetal birth weight in animals receiving arginine and L-citrulline supplementation (101, 102). These preclinical data demonstrate the potential of arginine supplementation to improve birth outcomes in the context of low-protein diets observed in pregnant women in LMICs. Arginine supplementation in human pregnancy has not been evaluated in low-income settings; however, studies from high- and middle-income countries have reported benefits of arginine supplementation in pre-eclampsia, gestational hypertension, FGR, and preterm labor (Table 1).

Arginine supplementation in pre-eclampsia

Pre-eclampsia, similar to malaria in pregnancy, is characterized by endothelial dysfunction and placental insufficiency, and there is evidence for dysregulation of the arginine-NO pathway in its pathophysiology (43, 71, 73). An initial trial of arginine supplementation in pre-eclampsia was underpowered and low quality, although it reported reductions in maternal blood pressure (103). Better-designed, yet still generally underpowered randomized controlled trials (RCTs) followed. Two evaluated longer-term, low-dose arginine supplementation (3 g/d for 3 wk) in pre-eclampsia and observed significant improvements to maternal and fetal hemodynamics, as well as better fetal outcomes (107, 108). In contrast, 2 RCTs in pre-eclamptic women using short-term, moderate doses initiated later in pregnancy (e.g., 12 g/d for 5 d) observed no benefit (104, 105). These data suggest that short-term supplementation, especially later in pregnancy, is insufficient to improve maternal hemodynamics.

The aforementioned observations are consistent with human and preclinical data indicating that interventions to affect placental vascular insufficiency via the arginine-NO pathway need to be initiated early and continued over the course of pregnancy (118–120). In support of this hypothesis, the best-designed and largest RCTs to date examined lowdose long-term arginine supplementation in pre-eclampsia (3 g or 6.6 g/d), initiated early in pregnancy (\sim 20 weeks of gestation) (76, 106). These studies reported arginine supplementation was associated with persistent decreases in maternal blood pressure and resulted in significantly improved fetal and maternal outcomes in pregnancies at risk of pre-eclampsia (76, 106).

 TABLE 1
 Characteristics of clinical studies providing L-arginine during pregnancy

Study (ref.)	Country	Dose (g/d)	Delivery form	Duration	Group	Treatment groups (n)	Key findings
PE studies Facchinetti et al. (103)	Italy	30	<u>``</u>	Single dose	n = 29	PE: uncomplicated pregnancy (12), PE (17)	 L-arginine-induced reduction in SBP and DBP in both groups, greater decrease of DBP in women with PE Increased serum L-citrulline during L-arginine treatment Total L-citrulline production inversely related begins a hood by programment accordated with high outcomes
Hladunewich et al. (104)	United States	14 (oral) or 30 (iv.)	Oral (i.v. if could not take oral)	Varied by participant: ≥3 d postpartum (some began predelivery)	n = 67 (n = 45 received treatment)	PE: L-arginine (22) or placebo (23), and healthy gravid controls (22)	 Increased baseline ADMA, cGMP, and endothelin in PE compared with healthy controls Increased postpartum serum L-arginine in the L-arginine treatment group No changes in postpartum serum NO, endothelin-1, cGMP, ADMA No change in postpartum glomerular filtration rate, blood pressure professions:
Staff et al. (105)	Norway	12	Oral	≥5 d	n = 30	PE: L-arginine (15) or	No difference in Japans van delivery or mean birth weight
Vadillo-Ortega et al. (76)	Mexico	<u>0</u>	Oral	Duration of pregnancy after enrollment (14–32 weeks of gestation)	n = 672	PErworen with previous PE or high risk; placebo (222), L-arginine + antioxidant vitamins (228), antioxidants alone (222)	Plasma L-arginine (before treatment) lower in women who later developed PE Reduced incidence of PE in L-arginine–treated group compared with placebo Benefit of L-arginine and antioxidant treatment over treatment with antioxidant vitamins alone Increased circulating L-arginine and decreased SBP and DBP in L-arginine + vitamins group Reduced risk of PTB in women who received L-arginine
Camarena Pulido et al. (106)	Mexico	М	Oral (capsules)	Duration of pregnancy after enrollment (20 weeks of gestation)	n = 96	High risk of PE: L-arginine treated (49) or placebo (47)	Increased incidence of PE, specifically severe PE, in placebo-treated women Increased birth weight and decreased PTB in Larginine—treated group
Rytlewski et al. (107)	Poland	m	Oral	3 wk	n = 61	PE: L-arginine (30) or placebo (31)	Lower SBP, DBP, and mean arterial blood pressure Lower SBP, DBP, and mean arterial blood pressure in L-arginine—treated group L-arginine increased 24-h urinary excretion of NOx and mean plasma L-citrulline No change in plasma L-arginine, L-ornithine, ADMA, SDMA, orl -NMMA
Rytlewski et al. (108)	Poland	m	Oral	Duration of pregnancy after enrollment (average enrollment at 29 weeks of gestation)	n = 61	PE: L-arginine (30) or placebo (31)	Langinine reduced pulsatility indexes in umbilical artery and increased pulsatility indexes in the middle cerebral artery, as well as increased cerebro-placental ratio values (MCA/UA) Decreased antihypertensive dosage in patients receiving Larginine Lower rates of FGR, increased latency to delivery, and higher Apgar scores in Larginine group

TABLE 1 (Continued)

Study (ref.)	Country	Dose (g/d)	Delivery form	Duration	Group	Treatment groups (n)	Key findings
Gestational hypertension studies Neri et al. (109)	Italy	20	<u>;;</u>	Single dose	n=15	Mild to moderate	No impact on fetal movements or fetal heart rate Action accining transmission and rate and representations are actions at the representation and representations are actions and repr
Neri et al. (110)	Italy	70	ž	2 9	n = 123	gestational hypertension: Gestational hypertension: L-arginine (62) or placebo (61)	 Acute algume treatment reduced flagentar for and Darwell Reduced SBP and DBP by both placebo and Larginine, with greater decrease in L-arginine group Improved performance of L-arginine (impact on blood pressure) in a subgroup analysis of women not receiving antihyoperpassives.
Facchinetti et al. (111)	Italy	20 (i.v.), 4 (oral)	iv. + oral	5 d i.v., then 2 wk oral	n = 74	Gestational hypertension with or without proteinuria: placebo (35) or L-arginine (39)	 Increased latency (to delivery) in L-arginine–treated group Reduced SBP and DBP 6 d post-ix. treatment L-arginine–associated trend towards increased latency, attenuated evolution of PE, and reduced low birth weight in subset of patients without promeinuria
Neri et al. (112) Advaca birth outroma studies	Italy	4	Oral	Duration of pregnancy after enrollment (< 16 weeks of gestation)	n = 79	Mild chronic hypertension: L-arginine (39) or placebo (40)	No difference in SBP or DBP, but fewer women in the L-arginine—treated group were given antihypertensive medications Increased gestational age, birth weight, and trend towards reduced neonatal complications in L-arginine group
Neri et al. (113)	Italy	30	<u>×</u>	Single dose	n = 27	FGR. AGA (9), and FGR with (9) or without (9) increased utero-placental resistance	 No hemodynamic changes in utero-umbilical circulation Larginine-induced decrease in resistance (nonplacental side) in FGR pregnancies with increased baseline resistance Increased plasma NOx and growth hormone
Winer et al. (114)	France	41	Oral	Duration of pregnancy after enrollment (24–32 weeks of gestation)	n = 43	Severe FGR: L-arginine (21) and placebo (22)	One change in birth weight or maternal or neonatal characteristics No impact on outcomes in pregnancies with severe vascular growth restriction No difference in severe NOx
Sieroszewski et al. (115)	Poland	ю	Oral	20 d	n = 108	FGR: L-arginine (78) or no treatment (30)	A connection of the second of the secon
Singh et al. (116)	India	м	Oral	21 d	n = 120	FGR: women with FGR (60; 30 received L-arginine, 30 placebo) or AGA fetal growth (60)	• Increased arternation was in a first than in AGA • Increased serum NO in Larginine—treated FGR • Increased serum NO in Larginine—treated FGR • Reduced umbilical artery resistance in Larginine—treated FGR group • Larginine increased live births, birth weight, and reduced neonatal complications (nonsticant)
Rytlewski et al. (117)	Poland	м	Oral	Varied by participant (average of 3 wk)	n = 45	Preterm labor: L-arginine (25) or placebo (20)	L-arginine reduced pulsatility indexes in umbilical artery and increased pulsatility indexes in the middle cerebral artery, as well as increased cerebro-placental ratio values (MCA/UA) No change in markers from NO pathway (plasma L-arginine, L-citrulline, L-ornithine, NOx) No change in pregnancy outcomes

¹Only original human studies of arginine supplementation in pregnancy are included. Animal studies and systematic reviews/meta-analyses are not included. ADMA, asymmetric dimethylarginine; AGA, appropriate for gestational age; cGMP, cyclic guanosine monophosphate; DBP, diastolic blood pressure; FGR, fetal growth restriction; L-NG-monomethyl arginine; MCA/UA, middle cerebral artery/umbilical artery; NOx, nitrate/nitrite; PE, pre-eclampsia; PTB, preterm birth; SBP, systolic blood pressure; SDMA, symmetric dimethylarginine.

Arginine supplementation in gestational hypertension

Four published RCTs have investigated arginine supplementation in the context of gestational hypertension (Table 1). The first 2 studies assessed the acute effects of large doses (20 g intravenously once, or for 5 d) relatively late in pregnancy (109, 110). Whereas 1 study was small (n = 15) (109), the other was a larger (n = 123) multicenter RCT (110). Both studies reported that parenteral arginine decreased maternal blood pressure but neither reported on pregnancy outcomes. In the absence of evidence of improved clinical outcomes, and given the challenges of long-term parenteral arginine administration, it is unclear whether this is a feasible therapeutic approach. In contrast, 2 subsequent RCTs ($\sim n = 70$) using longer-term, low-dose oral arginine (4 g/d) reported reduced maternal blood pressure (and/or reduced antihypertensive use) and improved pregnancy outcomes (111, 112).

Two meta-analyses of studies of arginine supplementation in the context of gestational hypertension and pre-eclampsia reported significantly decreased maternal diastolic blood pressure and extended length of pregnancy (121, 122). Collectively, the existing data support the use of longer-term low-dose arginine supplementation as the regimen most likely to improve maternal and fetal outcomes in hypertensive disorders of pregnancy.

Arginine supplementation and adverse birth outcomes

Many of the trials summarized in Table 1 evaluated adverse birth outcomes as either a primary or a secondary outcome (Table 1). Two trials that gave short-term arginine supplementation to women with pre-eclampsia reported no difference in birth outcomes between the 2 trial arms (104, 105). As above, these data are consistent with the hypothesis that short-term arginine supplementation initiated later in pregnancy is insufficient to reverse underlying vascular pathobiology. In contrast, the RCTs that used low-dose arginine supplementation early and continued it across pregnancy reported increased birth weight, length of gestation, and/or reduced rates of PTB (76, 106, 108, 111, 112).

Additional studies investigated arginine supplementation in the context of pregnancies complicated by FGR and preterm labor (Table 1). An initial report of arginine to treat FGR used a single arginine infusion (30 g intravenously) and reported improvements to utero-placental circulation (113). However, this trial was underpowered (n=27), uncontrolled, and relied on subgroup analyses, limiting its conclusions (113). Two larger RCTs of low-dose, long-term arginine supplementation (3 g/d for 3 wk) in pregnancies complicated by FGR reported improved neonatal outcomes and significantly increased fetal weight gain (115, 116).

Two trials of arginine supplementation in pregnancies with threatened adverse birth outcomes reported no benefit. One well-designed but small (n=43) multicenter RCT reported no difference in outcome in women with severe vascular FGR receiving either arginine (14 g/d) or placebo

(114). In another small trial of women presenting in preterm labor (n = 45), low-dose arginine supplementation (3 g/d) improved maternal and fetal hemodynamics but did not significantly improve birth outcomes (117). Both studies tested prolonged treatment with low dosage (114, 117); however, they enrolled severe cases of FGR or threatened PTB. Together these data indicate that arginine supplementation alone may not be sufficient to reverse advanced pathology including existing preterm labor or severe vascular FGR.

Summary and future directions for arginine supplementation studies

In summary, trials that reported significantly improved birth outcomes in women who received arginine compared with placebo (76, 106, 108, 111, 112, 115) used a prolonged, oral low-dose (3-7 g/d) regimen. Whereas, placebo-controlled trials that reported no difference in birth outcomes were in cases of advanced pathology (i.e., severe vascular FGR or preterm labor) (114, 117) or short-term treatment strategies initiated later in pregnancy (104, 105). To date, the largest (n = 672) well-designed RCT provided the strongest evidence that early initiation of low-dose arginine supplementation may improve pregnancy outcomes (76). In this study, women who began oral arginine supplementation (6.6 g/d) before 24 weeks of gestation had reduced rates of pre-eclampsia compared with placebo controls. Biologically, early and prolonged arginine dosing over pregnancy aligns with the important role of the arginine-NO biosynthetic pathway in regulating the critical processes of angiogenesis and vasculogenesis required for normal placental vascular development and healthy pregnancy outcomes. Once disrupted, these developmental pathways are unlikely to be reversed with late or single-dose approaches. Collectively the published human data support the hypothesis that supplementation with a daily physiological dose of arginine mimicking regular dietary intake (rather than a supraphysiological one-time dose) is beneficial to pregnancy outcomes in at-risk pregnancies.

Although mild gastrointestinal upsets such as dyspepsia were reported in large arginine supplementation trials in pregnancy (76, 106), these adverse events did not lead to participants discontinuing involvement in the studies. No study to date has reported any severe adverse events attributed to arginine supplementation in pregnancy. A limitation of the existing body of evidence is that most studies, with the exception of one (76), drew conclusions from relatively small sample sizes. Additional large RCTs, powered to investigate the impact of arginine on birth outcomes including FGR and PTB in at-risk pregnancies such as those in LMICs, are needed. Despite evidence that coadministration of citrulline and arginine may be a more effective strategy to increase bioavailable arginine and improve pregnancy outcomes in preclinical models (101, 102), no published human studies to date have examined the impact of supplementation with citrulline or a combination of arginine and citrulline on birth outcomes.

Dietary Arginine Intake and Impact on Birth Outcomes in Resource-Constrained Settings

Most research on arginine supplementation has been conducted in high-resource settings. The burdens of adverse birth outcomes, maternal malnutrition, and infections (i.e., malaria) that deplete arginine are greatest in LMICs, and therefore the benefit of arginine supplementation would be expected to be the largest in LMICs. There is preliminary evidence for an association between arginine and adverse birth outcomes in LMICs, but no existing studies have directly assessed arginine supplementation in LMICs. A recent study of >7000 Tanzanian pregnant women reported reduced risk of PTB in women with higher dietary arginine intake (123). Pan-African population-based analyses indicate dietary arginine deficiencies in resource-constrained settings (12) and therefore a greater risk of lower NO bioavailability during placental development. However, there is a need for RCTs in LMICs to address the hypothesis that arginine supplementation in pregnancy would improve birth outcomes in these settings.

Many LMICs (especially in sub-Saharan Africa, South Asia, and Southeast Asia) are endemic for malaria. In these regions malaria in pregnancy is a major contributor to poor birth outcomes, and a growing body of evidence is documenting the impact of malaria infection on bioavailable arginine and NO. Reduced circulating arginine concentrations and NO bioavailability, and a lower ratio of arginine to ADMA (a measure of arginine bioavailability) are well described in the context of malaria infection and correlate with endothelial dysfunction and disease severity (124–132). In these studies, recovery from severe malaria was associated with increases in circulating arginine (127), and intravenous arginine treatment of patients with severe malaria improved endothelial function and increased exhaled NO (126). Malaria-associated reductions in arginine and NO bioavailability may be attributed to increased circulating arginase and NO-scavenging cellfree hemoglobin released during malaria-induced hemolysis (86).

A study of Malawian women showed that malaria infection during pregnancy increased ADMA and symmetric dimethylarginine, and decreased bioavailable arginine (118). Malaria infection in early pregnancy was associated with reduced arginine concentrations and higher ADMA concentrations across pregnancy, and elevated ADMA early in pregnancy increased the relative risk of a small-forgestational-age delivery. Furthermore, measures of better maternal nutrition (e.g., higher midupper arm circumference and hemoglobin) were associated with increased circulating arginine, decreased ADMA, and increased ratio of arginine to ADMA. Although no human trials have yet reported arginine supplementation in the context of malaria in pregnancy, arginine supplementation in a preclinical mouse model of malaria in pregnancy enhanced placental vascular development and improved fetal weight and viability (118). These studies suggest that in settings with high rates of maternal malnutrition and malaria infection, maternal hypoarginemia could be a preventable and modifiable risk factor contributing to adverse birth outcomes such as PTB and FGR.

Conclusion

The burden of adverse birth outcomes in resourceconstrained settings is disproportionately high (1). New interventions appropriate for implementation in LMICs are needed to reduce poor birth outcomes and enable all children to survive and thrive. Population-level data of dietary arginine intake suggest that many women in LMICs are not receiving adequate dietary arginine required for successful pregnancy outcomes. Many LMICs are also malaria-endemic, an infection that has been shown to further deplete circulating arginine in pregnancy and contribute to placental insufficiency and adverse birth outcomes. Arginine deficiency in LMICs represents a modifiable risk factor to promote healthy birth outcomes. Based on the characteristics of food consumption in LMICs (e.g., food insecurity, growing food at home) and the large amount of daily arginine required in pregnancy (4-5 g/d; possibly more in malaria-endemic settings), we hypothesize that supplementation would represent an effective intervention strategy. We further propose that a combination supplement containing arginine and L-citrulline may most efficiently increase bioavailable arginine and NO in pregnant women.

Considering the preclinical and clinical evidence presented in this Perspective, we propose that a multisite RCT that evaluates ~7 g/d each of supplemental arginine and citrulline compared with placebo given to pregnant women in malaria-endemic areas, beginning early in pregnancy (<20 weeks of gestation) and continued over the course of pregnancy, would be an important next step to evaluate the potential of arginine to improve birth outcomes in LMICs.

The majority of preterm deliveries and adverse birth outcomes, including FGR, occur where women do not have access to adequate nutritional support. Preventing adverse birth outcomes (e.g., PTB, FGR) is one of the most costeffective strategies to improve global health and promote healthy life trajectories for young children. Confirming the safety, efficacy, cost-effectiveness, and scalability of arginine supplementation during pregnancy in reducing adverse birth outcomes would represent a major advance towards an affordable, culturally acceptable, and potentially life-saving intervention suitable for implementation and scale-up in low-resource settings.

Acknowledgments

The authors' contributions were as follows—CRM, ALC, WWF, and KCK: project conception and oversight; J-ABB, AMW, and CRM: research, analysis, and writing; and all authors: read and approved the final manuscript.

References

1. Blencowe H, Cousens S, Oestergaard MZ, Chou D, Moller A-B, Narwal R, Adler A, Vera Garcia C, Rohde S, Say L, et al. National, regional, and worldwide estimates of preterm birth rates in the year 2010 with

- time trends since 1990 for selected countries: a systematic analysis and implications. Lancet 2012;379(9832):2162–72.
- Lozano R, Naghavi M, Foreman K, Lim S, Shibuya K, Aboyans V, Abraham J, Adair T, Aggarwal R, Ahn SY, et al. Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the Global Burden of Disease Study 2010. Lancet 2012;380(9859):2095–128.
- Lawn JE, Davidge R, Paul VK, von Xylander S, de Graft Johnson J, Costello A, Kinney MV, Segre J, Molyneux L. Born too soon: care for the preterm baby. Reprod Health 2013;10(Suppl 1):S5.
- Black RE, Alderman H, Bhutta ZA, Gillespie S, Haddad L, Horton S, Lartey A, Mannar V, Ruel M, Victora CG, et al. Maternal and child nutrition: building momentum for impact. Lancet 2013;382(9890):372–5.
- Institute of Medicine. Preterm birth: causes, consequences and prevention [Internet]. Washington (DC): National Academies Press; 2007 [cited 16 Jan, 2019]. Available from: http://www.nap.edu/catalog/ 11622
- Mwaniki MK, Atieno M, Lawn JE, Newton CR. Long-term neurodevelopmental outcomes after intrauterine and neonatal insults: a systematic review. Lancet 2012;379(9814):445–52.
- Lawn JE, Kinney MV, Belizan JM, Mason E, McDougall L, Larson J, Lackritz E, Friberg IK, Howson CP. Born Too Soon: accelerating actions for prevention and care of 15 million newborns born too soon. Reprod Health 2013;10(Suppl 1):S6.
- Black RE, Victora CG, Walker SP, Bhutta ZA, Christian P, De Onis M, Ezzati M, Grantham-McGregor S, Katz J, Martorell R, et al. Maternal and child undernutrition and overweight in low-income and middle-income countries. Lancet 2013;382(9890): 427–51
- Christian P, Mullany LC, Hurley KM, Katz J, Black RE. Nutrition and maternal, neonatal, and child health. Semin Perinatol 2015;39(5): 361–72
- Victora CG, Adair L, Fall C, Hallal PC, Martorell R, Richter L, Sachdev HS. Maternal and child undernutrition: consequences for adult health and human capital. Lancet 2008;371(9609):340–57.
- FAO. The State of Food Security and Nutrition in the World 2017: Building Resilience for Peace and Food Security [Internet]. Rome: FAO; 2017 [cited 16 Jan, 2019]. Available from: www.fao.org/3/a-17695e.pdf
- Schönfeldt HC, Gibson Hall N. Dietary protein quality and malnutrition in Africa. Br J Nutr 2012;108(Suppl 2):S69–76.
- Semba RD, Shardell M, Sakr Ashour FA, Moaddel R, Trehan I, Maleta KM, Ordiz MI, Kraemer K, Khadeer MA, Ferrucci L, et al. Child stunting is associated with low circulating essential amino acids. EBioMedicine 2016;6:246–52.
- 14. Bhutta ZA, Das JK, Rizvi A, Gaffey MF, Walker N, Horton S, Webb P, Lartey A, Black RE. Evidence-based interventions for improvement of maternal and child nutrition: what can be done and at what cost? Lancet 2013;382(9890):452–77.
- Ahmed T, Hossain M, Sanin KI. Global burden of maternal and child undernutrition and micronutrient deficiencies. Ann Nutr Metab 2012;61(Suppl 1):8–17.
- Kelly A, Kevany J, de Onis M, Shah PM. A WHO collaborative study of maternal anthropometry and pregnancy outcomes. Int J Gynecol Obstet 1997;53(3):219–33.
- Sinha S, Patro N, Patro IK. Maternal protein malnutrition: current and future perspectives of spirulina supplementation in neuroprotection. Front Neurosci 2018;12:966.
- Imdad A, Bhutta ZA. Maternal nutrition and birth outcomes: effect of balanced protein-energy supplementation. Paediatr Perinat Epidemiol 2012;26(Suppl 1):178–90.
- Kramer MS, Kakuma R. Energy and protein intake in pregnancy. Cochrane Database Syst Rev 2003;(4):CD000032.
- Dellicour S, Tatem AJ, Guerra CA, Snow RW, ter Kuile FO. Quantifying the number of pregnancies at risk of malaria in 2007: a demographic study. PLoS Med 2010;7:e1000221.

- Desai M, ter Kuile FO, Nosten F, McGready R, Asamoa K, Brabin B, Newman RD. Epidemiology and burden of malaria in pregnancy. Lancet Infect Dis 2007;7(2):93–104.
- 22. Walker PGT, Floyd J, ter Kuile F, Cairns M. Estimated impact on birth weight of scaling up intermittent preventive treatment of malaria in pregnancy given sulphadoxine-pyrimethamine resistance in Africa: a mathematical model. PLoS Med 2017;14: e1002243
- Unger HW, Ashorn P, Cates JE, Dewey KG, Rogerson SJ. Undernutrition and malaria in pregnancy – a dangerous dyad? BMC Med 2016;14(1):142.
- 24. Cates JE, Unger HW, Briand V, Fievet N, Valea I, Tinto H, D'Alessandro U, Landis SH, Adu-Afarwuah S, Dewey KG, et al. Malaria, malnutrition, and birthweight: a meta-analysis using individual participant data. PLoS Med 2017;14(2):e1002373.
- 25. Wu G, Bazer FW, Cudd TA, Meininger CJ, Spencer TE. Maternal nutrition and fetal development. J Nutr 2004;134(9):2169–72.
- 26. Geva E, Ginzinger DG, Zaloudek CJ, Moore DH, Byrne A, Jaffe RB. Human placental vascular development: vasculogenic and angiogenic (branching and nonbranching) transformation is regulated by vascular endothelial growth factor-A, angiopoietin-1, and angiopoietin-2. J Clin Endocrinol Metab 2002;87(9):4213–24.
- Reynolds LP, Borowicz PP, Caton JS, Vonnahme KA, Luther JS, Buchanan DS, Hafez SA, Grazul-Bilska AT, Redmer DA. Uteroplacental vascular development and placental function: an update. Int J Dev Biol 2010;54(2–3):355–66.
- Vedmedovska N, Rezeberga D, Teibe U, Melderis I, Donders GGG. Placental pathology in fetal growth restriction. Eur J Obstet Gynecol Reprod Biol 2011;155(1):36–40.
- Triunfo S, Lobmaier S, Parra-Saavedra M, Crovetto F, Peguero A, Nadal A, Gratacos E, Figueras F. Angiogenic factors at diagnosis of late-onset small-for-gestational age and histological placental underperfusion. Placenta 2014;35(6):398–403.
- Noris M, Perico N, Remuzzi G. Mechanisms of disease: pre-eclampsia. Nat Clin Pract Nephrol 2005;1(2):98–114.
- 31. Ferrazzi E, Rigano S, Bozzo M, Bellotti M, Giovannini N, Galan H, Battaglia FC. Umbilical vein blood flow in growth-restricted fetuses. Ultrasound Obstet Gynecol 2000;16(5):432–8.
- 32. Konje JC, Howarth ES, Kaufmann P, Taylor DJ. Longitudinal quantification of uterine artery blood volume flow changes during gestation in pregnancies complicated by intrauterine growth restriction. BJOG 2003;110(3):301–5.
- Wang Y, Zhao S. Vascular Biology of the Placenta. San Rafael, CA: Morgan & Claypool Life Sciences; 2010.
- 34. Conroy AL, Silver KL, Zhong K, Rennie M, Ward P, Sarma JV, Molyneux ME, Sled J, Fletcher JF, Rogerson S, et al. Complement activation and the resulting placental vascular insufficiency drives fetal growth restriction associated with placental malaria. Cell Host Microbe 2013;13(2):215–26.
- 35. Wu G, Morris SM. Arginine metabolism: nitric oxide and beyond. Biochem J 1998;336(Pt 1):1–17.
- Böger RH, Bode-Böger SM. The clinical pharmacology of L-arginine. Annu Rev Pharmacol Toxicol 2001;41:79–99.
- Kielstein JT, Fliser D, Veldink H. Asymmetric dimethylarginine and symmetric dimethylarginine: axis of evil or useful alliance? Semin Dial 2009;22(4):346–50.
- Krause BJ, Hanson MA, Casanello P. Role of nitric oxide in placental vascular development and function. Placenta 2011;32(11):797–805.
- Foster MW, McMahon TJ, Stamler JS. S-nitrosylation in health and disease. Trends Mol Med 2003;9(4):160–8.
- Elphinstone RE, Besla R, Shikatani EA, Lu Z, Hausladen A, Davies M, Robbins CS, Husain M, Stamler JS, Kain KC. Snitrosoglutathione reductase deficiency confers improved survival and neurological outcome in experimental cerebral malaria. Infect Immun 2017;85(9):e00371–17.
- 41. De Falco S. The discovery of placenta growth factor and its biological activity. Exp Mol Med 2012;44(1):1–9.

- 42. Rossmanith WG, Hoffmeister U, Wolfahrt S, Kleine B, McLean M, Jacobs RA, Grossman AB. Expression and functional analysis of endothelial nitric oxide synthase (eNOS) in human placenta. Mol Hum Reprod 1999;5(5):487-94.
- 43. Böger RH, Diemert A, Schwedhelm E, Lüneburg N, Maas R, Hecher K. The role of nitric oxide synthase inhibition by asymmetric dimethylarginine in the pathophysiology of preeclampsia. Gynecol Obstet Invest 2010;69(1):1-13.
- 44. Böger RH, Bode-Böger SM, Frölich JC. The L-arginine-nitric oxide pathway: role in atherosclerosis and therapeutic implications. Atherosclerosis 1996;127(1):1–11.
- 45. Dhanakoti SN, Brosnan JT, Herzberg GR, Brosnan ME. Renal arginine synthesis: studies in vitro and in vivo. Am J Physiol 1990;259(3 Pt
- 46. Luiking YC, Poeze M, Ramsay G, Deutz NE. Reduced citrulline production in sepsis is related to diminished de novo arginine and nitric oxide production. Am J Clin Nutr 2009;89(1):142-52.
- 47. Castillo L, Chapman TE, Sanchez M, Yu YM, Burke JF, Ajami AM, Vogt J, Young VR. Plasma arginine and citrulline kinetics in adults given adequate and arginine-free diets. Proc Natl Acad Sci U S A 1993;90(16):7749-53.
- 48. Kao CC, Hsu JW, Dwarkanath P, Karnes JM, Baker TM, Bohren KM, Badaloo A, Thame MM, Kurpad AV, Jahoor F. Indian women of childbearing age do not metabolically conserve arginine as do American and Jamaican women. J Nutr 2015;145(5):884-92.
- 49. Wu G, Bazer FW, Satterfield MC, Li X, Wang X, Johnson GA, Burghardt RC, Dai Z, Wang J, Wu Z. Impacts of arginine nutrition on embryonic and fetal development in mammals. Amino Acids 2013;45(2):241-56.
- 50. Igarashi K, Kashiwagi K. Modulation of cellular function by polyamines. Int J Biochem Cell Biol 2010;42(1):39-51.
- 51. Bazer FW, Burghardt RC, Johnson GA, Spencer TE, Wu G. Mechanisms for the establishment and maintenance of pregnancy: synergies from scientific collaborations. Biol Reprod 2018;99(1): 225-41.
- 52. Lefèvre PLC, Palin M-F, Murphy BD. Polyamines on the reproductive landscape. Endocr Rev 2011;32(5):694-712.
- 53. Harris LK, McCormick J, Cartwright JE, Whitley GSJ, Dash PR. Snitrosylation of proteins at the leading edge of migrating trophoblasts by inducible nitric oxide synthase promotes trophoblast invasion. Exp Cell Res 2008;314(8):1765-76.
- 54. Kaufmann P, Black S, Huppertz B. Endovascular trophoblast invasion: implications for the pathogenesis of intrauterine growth retardation and preeclampsia. Biol Reprod 2003;69(1):1-7.
- 55. Ahmed A, Dunk C, Kniss D, Wilkes M. Role of VEGF receptor-1 (Flt-1) in mediating calcium-dependent nitric oxide release and limiting DNA synthesis in human trophoblast cells. Lab Invest 1997;76(6): 779-91.
- 56. Dunk C, Shams M, Nijjar S, Rhaman M, Qiu Y, Bussolati B, Ahmed A. Angiopoietin-1 and angiopoietin-2 activate trophoblast tie-2 to promote growth and migration during placental development. Am J Pathol 2000;156(6):2185-99.
- 57. Dötsch J, Hogen N, Nyúl Z, Hänze J, Knerr I, Kirschbaum M, Rascher W. Increase of endothelial nitric oxide synthase and endothelin-1 mRNA expression in human placenta during gestation. Eur J Obstet Gynecol Reprod Biol 2001;97(2):163-7.
- 58. Fukumura D, Gohongi T, Kadambi A, Izumi Y, Ang J, Yun CO, Buerk DG, Huang PL, Jain RK. Predominant role of endothelial nitric oxide synthase in vascular endothelial growth factor-induced angiogenesis and vascular permeability. Proc Natl Acad Sci U S A 2001;98(5): 2604 - 9.
- 59. Groesch KA, Torry RJ, Wilber AC, Abrams R, Bieniarz A, Guilbert LJ, Torry DS. Nitric oxide generation affects pro- and anti-angiogenic growth factor expression in primary human trophoblast. Placenta 2011;32(12):926-31.
- 60. Ahmed A, Perkins J. Angiogenesis and intrauterine growth restriction. Best Pract Res Clin Obstet Gynaecol 2000;14(6):981-98.

- 61. Sandrim VC, Palei ACT, Metzger IF, Gomes VA, Cavalli RC, Tanus-Santos JE. Nitric oxide formation is inversely related to serum levels of antiangiogenic factors soluble fms-like tyrosine kinase-1 and soluble endogline in preeclampsia. Hypertension 2008;52(2):402-7.
- 62. Conde-Agudelo A, Papageorghiou A, Kennedy S, Villar J. Novel biomarkers for predicting intrauterine growth restriction: a systematic review and meta-analysis. BJOG 2013;120(6):681-94.
- 63. Darling AM, McDonald CR, Conroy AL, Hayford KT, Liles WC, Wang M, Aboud S, Urassa WS, Kain KC, Fawzi WW. Angiogenic and inflammatory biomarkers in midpregnancy and small-forgestational-age outcomes in Tanzania. Am J Obstet Gynecol 2014;211:
- 64. Gagnon R. Placental insufficiency and its consequences. Eur J Obstet Gynecol Reprod Biol 2003;110(Suppl 1):S99-107.
- 65. Helske S, Vuorela P, Carpén O, Hornig C, Weich H, Halmesmäki E. Expression of vascular endothelial growth factor receptors 1, 2 and 3 in placentas from normal and complicated pregnancies. Mol Hum Reprod 2001;7(2):205-10.
- 66. Levine RJ, Maynard SE, Qian C, Lim K-H, England LJ, Yu KF, Schisterman EF, Thadhani R, Sachs BP, Epstein FH, et al. Circulating angiogenic factors and the risk of preeclampsia. N Engl J Med 2004;350(7):672-83.
- 67. McDonald CR, Darling AM, Conroy AL, Tran V, Cabrera A, Liles WC, Wang M, Aboud S, Urassa W, Fawzi WW, et al. Inflammatory and angiogenic factors at mid-pregnancy are associated with spontaneous preterm birth in a cohort of Tanzanian women. PLoS One 2015;10(8):e0134619.
- 68. Morgan TK. Role of the placenta in preterm birth: a review. Am J Perinatol 2016;33(3):258-66.
- 69. Poon LCY, Zaragoza E, Akolekar R, Anagnostopoulos E, Nicolaides KH. Maternal serum placental growth factor (PIGF) in small for gestational age pregnancy at 11⁺⁰ to 13⁺⁶ weeks of gestation. Prenat Diagn 2008;28(12):1110-15.
- 70. Fickling SA, Williams D, Vallance P, Nussey SS, Whitley GS. Plasma concentrations of endogenous inhibitor of nitric oxide synthesis in normal pregnancy and pre-eclampsia. Lancet 1993;342(8865):
- 71. Kim YJ, Park HS, Lee HY, Ha EH, Suh SH, Oh SK, Yoo HS. Reduced Larginine level and decreased placental eNOS activity in preeclampsia. Placenta 2006;27(4-5):438-44.
- 72. Krause BJ, Carrasco-Wong I, Caniuguir A, Carvajal J, Farías M, Casanello P. Endothelial eNOS/arginase imbalance contributes to vascular dysfunction in IUGR umbilical and placental vessels. Placenta 2013;34(1):20-8.
- 73. Pettersson A, Hedner T, Milsom I. Increased circulating concentrations of asymmetric dimethyl arginine (ADMA), an endogenous inhibitor of nitric oxide synthesis, in preeclampsia. Acta Obstet Gynecol Scand 1998;77(8):808-13.
- 74. Goodrum LA, Saade GR, Belfort MA, Moise KJJ, Jahoor F. Arginine flux and nitric oxide production during human pregnancy and postpartum. J Soc Gynecol Investig 2003;10(7):400-5.
- 75. Holden DP, Fickling SA, Whitley GS, Nussey SS. Plasma concentrations of asymmetric dimethylarginine, a natural inhibitor of nitric oxide synthase, in normal pregnancy and preeclampsia. Am J Obstet Gynecol 1998;178(3):551-6.
- 76. Vadillo-Ortega F, Perichart-Perera O, Espino S, Avila-Vergara MA, Ibarra I, Ahued R, Godines M, Parry S, Macones G, Strauss JF. Effect of supplementation during pregnancy with L-arginine and antioxidant vitamins in medical food on pre-eclampsia in high risk population: randomised controlled trial. BMJ 2011;342:d2901.
- 77. Silk DB, Grimble GK, Rees RG. Protein digestion and amino acid and peptide absorption. Proc Nutr Soc 1985;44(1):63-72.
- 78. Böger RH. The pharmacodynamics of L-arginine. J Nutr 2007;137(6 Suppl 2):1650S-5S.
- 79. Hou Y, Yin Y, Wu G. Dietary essentiality of "nutritionally nonessential amino acids" for animals and humans. Exp Biol Med 2015;240(8):997-1007.

- McNeal CJ, Meininger CJ, Reddy D, Wilborn CD, Wu G. Safety and effectiveness of arginine in adults. J Nutr 2016;146(12): 2587S-93S.
- Wu Z, Hou Y, Hu S, Bazer FW, Meininger CJ, McNeal CJ, Wu G. Catabolism and safety of supplemental L-arginine in animals. Amino Acids 2016;48(7):1541–52.
- FDA. Guidance for Industry: Estimating the Maximum Safe Starting dose in Initial Clinical Trials for Therapeutics in Adult Healthy Volunteers [Internet]. FDA; 2005 [cited 16 Jan, 2019]. Available from: https://www.fda.gov/downloads/Drugs/Guidances/UCM078932.pdf.
- Shao A, Hathcock JN. Risk assessment for the amino acids taurine, L-glutamine and L-arginine. Regul Toxicol Pharmacol 2008;50(3): 376–99
- McNeal CJ, Meininger CJ, Wilborn CD, Tekwe CD, Wu G. Safety of dietary supplementation with arginine in adult humans. Amino Acids 2018;50(9):1215–29.
- 85. Institute of Medicine. Dietary Reference Intakes for Energy, Carbohydrate, Fiber, Fat, Fatty Acids, Cholesterol, Protein, and Amino Acids [Internet]. Washington (DC): National Academies Press; 2005 [cited 9 Mar, 2017]. Available from: http://www.nap.edu/catalog/10490.
- Weinberg JB, Lopansri BK, Mwaikambo ED, Granger DL. Arginine, nitric oxide, carbon monoxide, and endothelial function in severe malaria. Curr Opin Infect Dis 2008;21(5):468–75.
- Allen LH, De Benoist B, Dary O, Hurrell R. Guidelines on Food Fortification with Micronutrients. Geneva: World Health Organization; 2006.
- 88. Cox SE, Ellins EA, Marealle AI, Newton CR, Soka D, Sasi P, Luca Di Tanna G, Johnson W, Makani J, Prentice AM, et al. Ready-to-use food supplement, with or without arginine and citrulline, with daily chloroquine in Tanzanian children with sickle-cell disease: a double-blind, random order crossover trial. Lancet Haematol 2018;5(4): e147–60
- 89. O'Sullivan D, Brosnan JT, Brosnan ME. Hepatic zonation of the catabolism of arginine and ornithine in the perfused rat liver. Biochem J 1998;330(Pt 2):627–32.
- Wu G, Meininger CJ. Arginine nutrition and cardiovascular function. J Nutr 2000;130(11):2626–9.
- Schwedhelm E, Maas R, Freese R, Jung D, Lukacs Z, Jambrecina A, Spickler W, Schulze F, Böger RH. Pharmacokinetic and pharmacodynamic properties of oral L-citrulline and L-arginine: impact on nitric oxide metabolism. Br J Clin Pharmacol 2008;65(1):51–9.
- Waugh WH, Daeschner CW, Files BA, McConnell ME, Strandjord SE.
 Oral citrulline as arginine precursor may be beneficial in sickle cell disease: early phase two results. J Natl Med Assoc 2001;93(10):363–71.
- 93. Suzuki T, Morita M, Hayashi T, Kamimura A. The effects on plasma L-arginine levels of combined oral L-citrulline and L-arginine supplementation in healthy males. Biosci Biotechnol Biochem 2017;81(2):372–5.
- Lin G, Wang X, Wu G, Feng C, Zhou H, Li D, Wang J. Improving amino acid nutrition to prevent intrauterine growth restriction in mammals. Amino Acids 2014;46(7):1605–23.
- Gao K, Jiang Z, Lin Y, Zheng C, Zhou G, Chen F, Yang L, Wu G. Dietary L-arginine supplementation enhances placental growth and reproductive performance in sows. Amino Acids 2012;42(6): 2207–14.
- 96. Li J, Xia H, Yao W, Wang T, Li J, Piao X, Thacker P, Wu G, Wang F. Effects of arginine supplementation during early gestation (day 1 to 30) on litter size and plasma metabolites in gilts and sows. J Anim Sci 2015;93(11):5291–303.
- Bérard J, Bee G. Effects of dietary L-arginine supplementation to gilts during early gestation on foetal survival, growth and myofiber formation. Animal 2010;4(10):1680-7.
- Lassala A, Bazer FW, Cudd TA, Datta S, Keisler DH, Satterfield MC, Spencer TE, Wu G. Parenteral administration of L-arginine enhances fetal survival and growth in sheep carrying multiple fetuses. J Nutr 2011;141(5):849–55.

- Mateo RD, Wu G, Bazer FW, Park JC, Shinzato I, Kim SW. Dietary Larginine supplementation enhances the reproductive performance of gilts. J Nutr 2007;137(3):652–6.
- 100. Wu G, Bazer FW, Burghardt RC, Johnson GA, Kim SW, Li XL, Satterfield MC, Spencer TE. Impacts of amino acid nutrition on pregnancy outcome in pigs: mechanisms and implications for swine production. J Anim Sci 2010;88(13 Suppl):E195–204.
- 101. Bourdon A, Parnet P, Nowak C, Tran N-T, Winer N, Darmaun D. L-citrulline supplementation enhances fetal growth and protein synthesis in rats with intrauterine growth restriction. J Nutr 2016;146(3): 532–41.
- 102. Tran NT, Amarger V, Bourdon A, Misbert E, Grit I, Winer N, Darmaun D. Maternal citrulline supplementation enhances placental function and fetal growth in a rat model of IUGR: involvement of insulin-like growth factor 2 and angiogenic factors. J Matern Fetal Neonatal Med 2017;30(16):1906–11.
- 103. Facchinetti F, Longo M, Piccinini F, Neri I, Volpe A. L-arginine infusion reduces blood pressure in preeclamptic women through nitric oxide release. J Soc Gynecol Investig 1999;6(4):202–7.
- 104. Hladunewich MA, Derby GC, Lafayette RA, Blouch KL, Druzin ML, Myers BD. Effect of L-arginine therapy on the glomerular injury of preeclampsia: a randomized controlled trial. Obstet Gynecol 2006;107(4):886–95.
- 105. Staff AC, Berge L, Haugen G, Lorentzen B, Mikkelsen B, Henriksen T. Dietary supplementation with L-arginine or placebo in women with pre-eclampsia. Acta Obstet Gynecol Scand 2004;83(1):103–7.
- 106. Camarena Pulido EE, García Benavides L, Panduro Barón JG, Pascoe Gonzalez S, Madrigal Saray AJ, García Padilla FE, Totsuka Sutto SE. Efficacy of L-arginine for preventing preeclampsia in high-risk pregnancies: a double-blind, randomized, clinical trial. Hypertens Pregnancy 2016;35(2):217–25.
- 107. Rytlewski K, Olszanecki R, Korbut R, Zdebski Z. Effects of prolonged oral supplementation with L-arginine on blood pressure and nitric oxide synthesis in preeclampsia. Eur J Clin Invest 2005;35(1):32–7.
- 108. Rytlewski K, Olszanecki R, Lauterbach R, Grzyb A, Basta A. Effects of oral L-arginine on the foetal condition and neonatal outcome in preeclampsia: a preliminary report. Basic Clin Pharmacol Toxicol 2006;99(2):146–52.
- Neri I, Blasi I, Facchinetti F. Effects of acute L-arginine infusion on non-stress test in hypertensive pregnant women. J Matern Fetal Neonatal Med 2004;16(1):23–6.
- 110. Neri I, Jasonni VM, Gori GF, Blasi I, Facchinetti F. Effect of L-arginine on blood pressure in pregnancy-induced hypertension: a randomized placebo-controlled trial. J Matern Fetal Neonatal Med 2006;19(5): 277–81.
- 111. Facchinetti F, Saade GR, Neri I, Pizzi C, Longo M, Volpe A. L-arginine supplementation in patients with gestational hypertension: a pilot study. Hypertens Pregnancy 2007;26(1):121–30.
- 112. Neri I, Monari F, Sgarbi L, Berardi A, Masellis G, Facchinetti F. L-arginine supplementation in women with chronic hypertension: impact on blood pressure and maternal and neonatal complications. J Matern Fetal Neonatal Med 2010;23(12):1456–60.
- 113. Neri I, Mazza V, Galassi MC, Volpe A, Facchinetti F. Effects of L-arginine on utero-placental circulation in growth-retarded fetuses. Acta Obstet Gynecol Scand 1996;75(3):208–12.
- 114. Winer N, Branger B, Azria E, Tsatsaris V, Philippe HJ, Roze JC, Descamps P, Boog G, Cynober L, Darmaun D. L-arginine treatment for severe vascular fetal intrauterine growth restriction: a randomized double-bind controlled trial. Clin Nutr 2009;28(3):243–8.
- 115. Sieroszewski P, Suzin J, Karowicz-Bilińska A. Ultrasound evaluation of intrauterine growth restriction therapy by a nitric oxide donor (Larginine). J Matern Fetal Neonatal Med 2004;15(6):363–6.
- 116. Singh S, Singh A, Sharma D, Singh A, Narula MK, Bhattacharjee J. Effect of L-arginine on nitric oxide levels in intrauterine growth restriction and its correlation with fetal outcome. Indian J Clin Biochem 2015;30(3):298–304.
- 117. Rytlewski K, Olszanecki R, Lauterbach R, Grzyb A, Kiec-Wilk B, Dembinska-Kiec A, Basta A. Effects of oral L-arginine on the pulsatility

- indices of umbilical artery and middle cerebral artery in preterm labor. Eur J Obstet Gynecol Reprod Biol 2008;138(1):23-8.
- 118. McDonald CR, Cahill LS, Gamble JL, Elphinstone R, Gazdzinski LM, Zhong KJY, Philson AC, Madanitsa M, Kalilani-Phiri L, Mwapasa V, et al. Malaria in pregnancy alters L-arginine bioavailability and placental vascular development. Sci Transl Med 2018;10(431):eaan6007.
- 119. Paauw ND, Terstappen F, Ganzevoort W, Joles JA, Gremmels H, Lely AT. Sildenafil during pregnancy: a preclinical meta-analysis on fetal growth and maternal blood pressure. Hypertension 2017;70(5):998-1006.
- 120. Roberge S, Nicolaides K, Demers S, Hyett J, Chaillet N, Bujold E. The role of aspirin dose on the prevention of preeclampsia and fetal growth restriction: systematic review and meta-analysis. Am J Obstet Gynecol 2017;216(2):110-20.e6.
- 121. Gui S, Jia J, Niu X, Bai Y, Zou H, Deng J, Zhou R. Arginine supplementation for improving maternal and neonatal outcomes in hypertensive disorder of pregnancy: a systematic review. J Renin Angiotensin Aldosterone Syst 2014;15(1):88-96.
- 122. Zhu Q, Yue X, Tian Q-Y, Saren G, Wu M-H, Zhang Y, Liu T-T. Effect of L-arginine supplementation on blood pressure in pregnant women: a meta-analysis of placebo-controlled trials. Hypertens Pregnancy 2013;32(1):32-41.
- 123. Darling AM, McDonald CR, Urassa WS, Kain KC, Mwiru RS, Fawzi WW. Maternal dietary L-arginine and adverse birth outcomes in Dar es Salaam, Tanzania. Am J Epidemiol 2017;186(5):603-11.
- 124. Anstey BNM, Weinberg JB, Hassanali MY, Mwaikambo ED, Manyenga D, Misukonis MA, Arnellefi DR, Hollis D, McDonald MI, II, Granger DL. Nitric oxide in Tanzanian children with malaria: inverse relationship between malaria severity and nitric oxide production/nitric oxide synthase type 2 expression. J Exp Med 1996;184(2):557-67.
- 125. Lopansri BK, Anstey NM, Weinberg JB, Stoddard GJ, Hobbs MR, Levesque MC, Mwaikambo ED, Granger DL. Low plasma arginine concentrations in children with cerebral malaria and decreased nitric oxide production. Lancet 2003;361(9358):676-8.

- 126. Yeo TW, Lampah DA, Gitawati R, Tjitra E, Kenangalem E, McNeil YR, Darcy CJ, Granger DL, Weinberg JB, Lopansri BK, et al. Impaired nitric oxide bioavailability and L-arginine reversible endothelial dysfunction in adults with falciparum malaria. J Exp Med 2007;204(11): 2693-704.
- 127. Yeo TW, Lampah DA, Gitawati R, Tjitra E, Kenangalem E, McNeil YR, Darcy CJ, Granger DL, Weinberg JB, Bert K, et al. Recovery of endothelial function in severe falciparum malaria: relationship with improvement in plasma L-arginine and blood lactate concentrations. J Infect Dis 2008;198(4):602-8.
- 128. Yeo TW, Lampah DA, Tjitra E, Gitawati R, Darcy CJ, Jones C, Kenangalem E, McNeil YR, Granger DL, Lopansri BK, et al. Increased asymmetric dimethylarginine in severe falciparum malaria: association with impaired nitric oxide bioavailability and fatal outcome. PLoS Pathog 2010;6(4):e1000868.
- 129. Chertow JH, Alkaitis MS, Nardone G, Ikeda AK, Cunnington AJ, Okebe J, Ebonyi AO, Njie M, Correa S, Jayasooriya S, et al. Plasmodium infection is associated with impaired hepatic dimethylarginine dimethylaminohydrolase activity and disruption of nitric oxide synthase inhibitor/substrate homeostasis. PLoS Pathog 2015;11(9):e1005119.
- 130. Weinberg JB, Yeo TW, Mukemba JP, Florence SM, Volkheimer AD, Wang H, Chen Y, Rubach M, Granger DL, Mwaikambo ED, et al. Dimethylarginines: endogenous inhibitors of nitric oxide synthesis in children with falciparum malaria. J Infect Dis 2014;210(6): 913-22.
- 131. Barber BE, William T, Grigg MJ, Parameswaran U, Piera KA, Yeo TW, Anstey NM. Asymmetric dimethylarginine in adult falciparum malaria: relationships with disease severity, antimalarial treatment, hemolysis, and inflammation. Open Forum Infect Dis 2016;3(1):ofw027.
- 132. Alkaitis MS, Wang H, Ikeda AK, Rowley CA, MacCormick IJC, Chertow JH, Billker O, Suffredini AF, Roberts DJ, Taylor TE, et al. Decreased rate of plasma arginine appearance in murine malaria may explain hypoargininemia in children with cerebral malaria. J Infect Dis 2016;214(12):1840-9.